RESEARCH ARTICLE

Lepidoptera demonstrate the relevance of Murray's Law to circulatory systems with tidal flow

Sandra R. Schachat^{1*} ^(D), C. Kevin Boyce¹, Jonathan L. Payne¹ and David Lentink^{2,3}

Abstract

Background: Murray's Law, which describes the branching architecture of bifurcating tubes, predicts the morphology of vessels in many amniotes and plants. Here, we use insects to explore the universality of Murray's Law and to evaluate its predictive power for the wing venation of Lepidoptera, one of the most diverse insect orders. Lepidoptera are particularly relevant to the universality of Murray's Law because their wing veins have tidal, or oscillatory, flow of air and hemolymph. We examined over one thousand wings representing 667 species of Lepidoptera.

Results: We found that veins with a diameter above approximately 50 microns conform to Murray's Law, with veins below 50 microns in diameter becoming less and less likely to conform to Murray's Law as they narrow. The minute veins that are most likely to deviate from Murray's Law are also the most likely to have atrophied, which prevents efficient fluid transport regardless of branching architecture. However, the veins of many taxa continue to branch distally to the areas where they atrophied, and these too conform to Murray's Law at larger diameters (e.g., Sesiidae). **Conclusions:** This finding suggests that conformity to Murray's Law in larger taxa may reflect requirements for structural support as much as fluid transport, or may indicate that selective pressures for fluid transport are stronger during the pupal stage—during wing development prior to vein atrophy—than the adult stage. Our results increase the taxonomic scope of Murray's Law and provide greater clarity about the relevance of body size.

Keywords: Fluid transport, Hemolypmh, Insect, Tracheae, Venation

Background

Biologists have long searched for universal "laws" that govern life on earth [1]. The wings of insects provide an amenable system for evaluating relevant biological laws because insects have the highest described species diversity of any group of animals [2] and their wings vary tremendously in terms of size, shape, and biomechanics [3]. Furthermore, insect wings serve a range of functions from thermoregulation to flight to sexual signaling, and the veins within insect wings serve sensory, structural, and circulatory roles [4].

The transport of air and hemolymph through the veins of insect wings is of particular relevance because the circulatory systems of animals and the vascular systems of plants present a rare opportunity to identify fundamental generalities that apply to more than one kingdom of multicellular life [5]. These efforts have expanded to include the rate of flow through vessels that bifurcate, such as an artery that branches into arterioles [6] or a vein in an insect wing. Flow through a bifurcating tube is often evaluated with the equation:

$$d_0^k = d_1^k + d_2^k + \dots + d_n^k$$

wherein a single tube with diameter d_0 branches into multiple tubes with diameters d_1 , d_2 , ..., d_n . Certain values of k, the "junction exponent" [7], represent distinct biophysical optima. When k = 2, the branching architecture conforms to "da Vinci's Law" [8] and flow velocity is conserved across the point of bifurcation; when k = 3, the branching architecture conforms to "Murray's Law" [9]

© The Author(s). 2021 **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated in a credit line to the data.





Open Access

^{*}Correspondence: sschachat@schmidtsciencefellows.org

¹Department of Geological Sciences, Stanford University, Stanford, USA Full list of author information is available at the end of the article

and transport capacity is conserved across the point of bifurcation; when k = 4, resistance to flow is conserved across the point of bifurcation [10].

At the biophysical optimum described by Murray's Law, a cost function is preserved before and after the vessel bifurcates [11]. There are two costs to moving fluid through a biological vessel. The first cost is the energy, or power, that fluid transport requires. This cost decreases as vessel radius (*r*) increases (power for steady flow $\sim 1/r^4$), assuming viscosity, kinetic, and gravity terms remain constant throughout the network. The second cost is the development and maintenance of the living cells that compose the wall of the vessel and the fluids therein. In contrast to the first cost, this second cost increases with the volume of the network (power associated with metabolism $\sim r^2 * length$ of the network). Vessel radius represents a tradeoff between these two costs and determines the total cost to the organism of the vessel network. Minimizing the two power terms leads to a definition of total flux proportional to r^3 . This definition can be applied to the special case of a bifurcation where preserving flux and minimizing the amount of power required results in $r_0^3 = r_1^3 + r_2^3$. Because we discuss our results in terms of vein diameter rather than radius, we employ an equivalent equation: $d_0^3 = d_1^3 + d_2^3$.

Studies of branching architecture have typically found support for Murray's Law [6]. Murray's Law was originally developed to describe blood flow in mammals, the system for which the greatest quantity of data is currently available [12, 13]. Murray's Law also describes fluid flow in various plants [14, 15] despite the vast differences between the circulatory systems of vertebrates and the vascular systems of plants. Vertebrates have a closed circulatory system of fixed volume on short timescales whereas xylem transport is subject to continual volume loss to evaporation, and vertebrate blood contains entire cells whereas plant vasculature transports water and solutes. One of the few commonalities that these transport systems share in plants and vertebrates is unidirectionality within individual conduits. Therefore, circulatory systems with tidal, or oscillatory, flow provide an opportunity to further test the universality of Murray's Law.

Although only a few species have been examined to date, the wing veins of obtectomeran Lepidoptera have been found to exhibit tidal flow [16, 17]. Air is displaced with each tracheal contraction, driving flow of both air and hemolymph and allowing more of the cross-sectional area of the vein to be occupied by hemolymph [16] (Fig. 1). This oscillation occurs many times per hour throughout the animal's adult lifespan [17].

The wing veins of Lepidoptera are far better suited for evaluation of Murray's Law than the wing veins of other insects for two additional reasons. First, the crosssectional shapes of wing veins in various insect orders



differ markedly from the circular to polygonal shape seen in both plants and vertebrates, which optimizes fluid flow by minimizing cross-sectional area in close proximity to the vein wall [4, 19–22]. The cross-sectional shapes documented in the wing veins of moths are far more circular than those documented in other insect orders [23, 24] and even approach the round shape of vertebrate and some plant veins [16]. This circular cross-sectional shape was recently shown not to be a requirement of Murray's Law [25] but nevertheless suggests optimization for fluid flow.

Second, the tracheae in insect wings typically occupy less than half of the available area within the veins but nevertheless occupy enough space to disrupt the patterns of hemolymph flow that would occur in their absence [4]. The dimensions of a typical vein cavity, therefore, merely exert an upper limit on the amount of space dedicated to the flow of air and hemolymph—these dimensions constrain but do not determine the transport capacity that Murray's Law describes. But in Lepidoptera, nearly all space within the vein cavity is dedicated to the flow of air when the tracheae expand, and more space becomes available for the flow of hemolymph when the tracheae contract [16] (Fig. 1).

The dimensions of the vein cavity determine the amount of space dedicated to the flow of both air and hemolymph. The space occupied by a trachea may well violate the assumptions of the Hagen–Poiseuille Law for the hemocoel of a lepidopteran wing vein, thus rendering Murray's Law inapplicable to the flow of hemolymph. Nonetheless, no such violations of the Hagen-Poiseuille Law are seen within the tracheae themselves. We therefore assume here that the Hagen-Poiseuille Law, and thus Murray's Law, apply to both the flow of air within the tracheae contained in wing veins and to the flow of hemolymph within wing veins that do not contain tracheae. A study of the atlas moth Attacus atlas L. (Saturniidae) found the trachea to occupy nearly the entirety of the vein cavity while expanded [18] such that the interior diameter of the vein determines conformity to Murray's Law for the flow of air. Critically, none of the mathematical derivations of Murray's Law [9, 25] assume the flow is from the parent to the child veins; the Law holds equally for reversed flow from the child veins to the parent vein during tidal flow.

Here we present an evaluation of Murray's Law for the wings of hundreds of species spanning the entire lepidopteran phylogeny. These wings vary in size, shape, and in the branching architecture of the venation (Figs. 2 and 3), permitting a nuanced perspective on the selective pressures that may underlie vein optimization for fluid transport. This study is both confirmatory and exploratory. It is confirmatory in testing the hypothesis that Murray's Law describes the geometry of vein branching patterns in Lepidoptera. It is also exploratory in assessing the extent to which conformity to Murray's Law varies as a function of taxonomic affiliation and parent vein diameter—relationships for which there is no pre-existing hypothesis to test.

Results

We found that Murray's Law consistently predicts branching architecture for bifurcations in which the parent vein has a diameter above 45–58 microns. For smaller veins, Murray's Law has less and less predictive power as diameter decreases. Bifurcation angle, unlike vein diameter, does not predict conformity to Murray's Law. A sensitivity analysis found that the images used are of sufficient pixel density for precise measurements of the diameters of vein cavities (Additional file 1: Figures S1, S2).

Murray's Law was evaluated here by solving for k in the equation outlined above. Because the veins of moth wings bifurcate into two child veins, the appropriate representation of the equation is: $d_0^k = d_1^k + d_2^k$. The bifurcations that most closely conform to Murray's Law are those for which k is closest to 3.

The values of k observed in Lepidoptera vary tremendously (Fig. 4; Additional file 2: Figure S3). Of the 2696 bifurcations examined, 127 have child veins that are both wider than the parent vein, yielding a negative value of k. Another 338 bifurcations have one child vein that is wider





than the parent vein and one that is narrower than the parent vein. Some bifurcations with this type of asymmetry yield values of k consistent with the various biophysical predictions outlined above, while others yield values of k below -100 or above 70. Both child veins are narrower than the parent vein, as predicted by Murray's Law and other theoretical optima, in 83% of the bifurcations examined.

But despite the sensitivity of k and the variability in the dataset, and regardless of how the dataset is processed whether or not negative values of k and asymmetrical bifurcations are included and whether or not k is truncated—a clear pattern emerges (Fig. 4a–d). The average deviation from Murray's Law (k = 3), defined simply as |k - 3|, decreases as the cross-sectional diameter of the parent vein (d_0) increases—until d_0 falls within the range of 45–58 microns (the 95% confidence interval spans diameters as low as 39 microns and as high as 65 microns). Beyond this point, deviation from Murray's Law remains low as d_0 continues to increase with the slope of this segment nearly equal to 0—ranging from - 0.0018 to - 0.0007—and the 95% confidence interval for the slope always includes 0. Visualization of our results in phylogenetic context demonstrates that our findings are not merely an artifact of phylogenetic autocorrelation (Fig. 5; Additional file 3: supplement 2).

Vein angle

In contrast to d_0 , the relationship between vein angle and k is not clear and may be an artifact of the phylogenetic relationships among the lineages sampled (Fig. 4h–n; Additional file 4: Figure S4). In the four datasets that contain all species, k conforms closely to Murray's Law in the bifurcations for where the two child veins connect at a very narrow angle, below approximately five degrees (Fig. 4h–k). k is more variable in the bifurcations with wider angles.



the mean deviation will not reach zero simply due to measurement error or any small deviation

The few bifurcations with angles greater than 40° tend to yield values of k that deviate strongly from Murray's Law. However, unlike d_0 , vein angle appears to have a relationship with k that is largely an artifact of the phylogenetic relationships among the taxa sampled, as shown by the stratification of superfamilies within higher grades and clades (Fig. 4l–n). Vein angle varies widely in smaller moths but does not exceed 30° in larger moths, further

supporting the notion that selective pressures increase with vein diameter.

The relationship between vein angle and fluid transport can also be evaluated by comparing the angle of each individual child vein to the angle of its parent vein, which can vary noticeably even when the angle between the child veins is small. Murray himself predicted that, when two child branches are of unequal diameter, the wider child



branch will occur at a lower angle to the parent branch [9, 41]. However, in Lepidoptera, there is no trend in the relationship between the difference in angle between the parent and child vein and the difference in diameter (Fig. 6).

Trichoptera: the sister-group to lepidoptera

The distribution of k among 82 bifurcations in the R vein on the wings of Trichoptera is very similar to the distribution seen in Lepidoptera (Fig. 7). However, the relationship between k and d_0 in Trichoptera is unclear (Fig. 7).

This relationship does not show the same sizedependence in Trichoptera as in Lepidoptera. When the same change-point analysis discussed above was implemented for Trichoptera, the 95% confidence interval recovered for the value of d_0 at the change point (23– 64 microns) was nearly identical to the entire range of d_0 in the dataset (23–67 microns). Furthermore, the 95% confidence intervals for the slope both before and after the change point include 0. It is unclear whether additional data, perhaps spanning a wider range of d_0 , would show a discernible pattern or a biological reason for this deviation.

Discussion

Murray's Law predicts branching architecture reasonably well in the wings of Lepidoptera. This finding of a shared biophysical optimum among the circulatory and vascular systems of Lepidoptera, vertebrates, and various plants suggests that Murray's Law is one of the few theoretical predictions that holds throughout the multicellular tree of life—at least at the diameters examined thus far.

The primary caveat to the applicability of Murray's Law in Lepidoptera is vein diameter: smaller moths, whose parent veins have diameters below 45-58 microns, are the least likely to conform to Murray's Law. The circulatory systems in the wings of these moths do not conform to a different biophysical optimum, and instead evince a far wider range of vales of k than is seen in larger moths. This suggests that circulatory systems in the wings of smaller moths are not under distinct selective pressures to those that determine vein morphology in larger moths, but instead are under weaker selective pressures.

The importance of hemolymph transport in the wings of Lepidoptera, particularly of small moths, is not entirely understood. In this order, hemocytes appear to occur at far greater abundances in larvae than in adults; although many hemocytes can be seen in the hemolymph circulating throughout the wing veins of the butterfly *Vanessa cardui* L. [17], data on hemocyte quantity in the wings of microlepidoptera are lacking, and the wing veins of many microlepidoptera may be too narrow to transport hemocytes. However, studies of Diptera show that hemolymph can continue to flow through wing veins even when the diameter of the vein cavity is too small to accommodate Schachat et al. BMC Biology (2021) 19:204







hemocytes [42, 43]. The importance of hemolymph transport in microlepidoptera, regardless of hemocyte content, is underscored by the widespread presence in Monotrysia of accessory pulsatile organs known as "wing hearts" [44]. Because these organs pump hemolymph into wing veins [45], their presence in small-bodied, early-diverging families such as Eriocraniidae and Lophocoronidae indicates that the hemolymph flow in these small moths is of sufficient physiological importance to require dedicated anatomical structures—although we do not have sufficient data from Lepidoptera to determine whether the cells that line vein walls [4, 46] conform to the assumptions about the cost function of Murray's Law.

The relaxation of selective pressures for animals with smaller body sizes has been noted in other taxa. When digits first originated in tetrapods their numbers varied tremendously but then stabilized very quickly [47]. Digit reduction and loss among extant taxa occurs frequently but predictably, in terms of both life history and the digits affected [48, 49]. This consistency often disappears, however, among the smallest-bodied salamander and frog genera, in which digits can become reduced and lost in a highly variable and seemingly random manner [50–52].

Lepidoptera are hardly the only clade that possesses a size threshold that separates strong and weak selective pressures. Weaker selective pressures at small body sizes are particularly common for gas exchange because diffusion becomes an adequate method of gas exchange upon sufficient reduction of body size [53].

In small moths, the decreased importance of the circulatory function of wing veins is readily apparent: veins regularly atrophy, both at the base and distally to the discal cell (Fig. 8). The wing veins of Lepidoptera are said to "atrophy" when they seemingly disappear from the wing [54], becoming visible only as "faint traces" if at all [44]. "Atrophied" veins were present in the developing wings of the pupa but are not intact in the adult wings. Although it cannot be stated with complete certainty that atrophied veins do not transport any fluid in the adult wing, their morphology is not optimized for fluid transport. The role of wing veins in circulation is limited even in small moths whose veins do not atrophy because much of the wing surface area used for flight derives from the wing fringe. The fringe consists only of scales, contains no living cells, occurs beyond the wing membrane, and therefore does not contain any veins. Furthermore, when the vein of a small moth bifurcates, the trachea within it often does not, and may not even extend far enough from the wing base to reach the point where the vein bifurcates [55, 56]. Furthermore, we note that many of the microlepidoptera examined here whose values of k deviate strongly from Murray's Law have atrophied veins and veins that only appear at the wing margin (Figs. 8 and 9).

Thus, conformity to Murray's Law in many lepidopteran taxa cannot be attributed solely to optimization for fluid transport in the adult wing. However, in addition to minimizing the pump work required to transport fluid, a network analysis by [57] has shown that vein networks adhering to Murray's Law have minimal total mass for a given flow transport rate. And as detailed in the supplemental material (Additional file 5), Murray's Law also describes a structural optimum for Lepidoptera whose wings are not corrugated. This provides a potential explanation for conformity to Murray's Law, and its size-dependence, in taxa such as Sesiidae whose veins atrophy before bifurcating. Alternatively, conformity to Murray's Law in moths with atrophied veins on the adult wing may be an artifact of optimization for fluid transport during the pupal



stage, before veins reach the atrophied state seen in the adult wing. Heightened conformity to Murray's Law during earlier stages of development has been observed in humans [58] and could be evaluated for Lepidoptera in future studies that examine pupal wings.

In the human circulatory system [11, 59–61], bifurcations follow da Vinci's Law more closely than Murray's Law at larger diameters. For all 455 bifurcations measured here in which d_0 is at least 100 microns—a threshold at which conformity to Murray's Law has definitely stabilized—the mean value of k is 2.91 (95% confidence interval: 2.75–3.09). Although k does not exactly equal 3, this value very nearly falls within the range observed in pigs [59] that provides strong empirical support for Murray's Law. However, for the 100 bifurcations with the largest values of d_0 , ranging from 360 to 932 microns, the mean value of k is 2.51 (95% confidence interval: 2.35-2.69). Future studies that examine venation in larger individuals belonging to the species Attacus atlas and in other large-bodied species such as Thysania agrippina (Cramer) (Erebidae) could provide greater insight into whether the wings of Lepidoptera conform more closely to da Vinci's law at larger vein diameters.

Conclusions

Lepidoptera are the most diverse taxon for which the applicability of Murray's Law has been examined. Our findings demonstrate the size-dependent relevance of Murray's Law to Lepidoptera, a conclusion that is robust to the evolutionary relatedness of the taxa sampled. Some of our findings, such as the potential relevance of da Vinci's Law to the largest vessels, are consistent with previous studies of both mammals and plants. However, our dataset also includes novel features such as the sharp decrease in conformity to Murray's Law in vessels below 50 microns and the relationship between Murray's Law and vessel size even for bifurcations that occur distally



to atrophied veins. Our findings highlight the universality of Murray's Law and raise questions about the selective pressures that may underlie its applicability.

Methods

The cross-sectional widths of the internal cavities of lepidopteran forewing veins were measured from microscope slides of "cleared" wings, from which the scales have been bleached and removed to facilitate visualization of veins. Measurements were taken from the radius vein (R), which in moths bifurcates into branches known as R_1 , R_2 , Rs_1 , Rs_2 , Rs_3 , and Rs_4 [24]. This vein was chosen because it typically bifurcates at least three times before connecting with branches of the medial vein via cross-veins. Forks in R were only measured if both of the resulting branches reach the margin of the wing, such that forks whose posterior branch forms a closed cell were not measured. Forks could not be measured when the branching point occurred immediately distal to the confluence of two veins. The points that were measured are illustrated on the examples provided in Figs. 2 and 3. Wings whose R vein bifurcates only twice were not included in this study unless those wings are less than six millimeters in length. All wing slides in the dataset were photographed in the collections where they are housed or were taken on loan and photographed at Stanford University with the exception of the ctenuchine and euchromiine wing slides, which were not available for loan and could only be measured from photographs.

Of the primary veins on the forewings of Lepidoptera, the subcosta branches in some homoneura [27, 62] and in Arrhenophanidae [63] but is unbranched in the vast majority of species, the cubitus posterior vein branches in exceedingly rare cases [26], and the anal veins are variable in microlepidoptera [27, 28, 62] and often lack any confluence or bifurcations in butterflies [35] and Macroheterocera [40, 64, 65]. This leaves the radius, medial, and cubitus anterior as the only veins with which Murray's Law can be evaluated across Lepidoptera. The radial sector typically branches into Rs1, Rs2, Rs3, and Rs4, permitting multiple measurements from the same vein. The branching patterns of the medial and cubitus anterior veins are far less consistent: the medial vein contains up to five branches [26], with three being the typical number outside Agathiphaga-with the crucial exception of Noctuoidea, the lepidopteran superfamily with the greatest described species diversity, in which countless "quadrifid" species have four branches of the medial vein [64]. Complicating matters further, free *M* and *CuA* veins both reach the base of the wing in various microlepidoptera but not in Obtectomera (Figs. 2 and 3). As noted by [66], this variation causes meaningful differences in hemolymph circulation.

Measurements

In each forewing, the *R* vein bifurcates at least once. Data were collected here for up to three points of bifurcation. At each point of bifurcation, the vein diameter was measured for all three components, the parent and two child branches. Along with a measurement of overall wing length, this resulted in four, seven, or ten measurements per wing, depending on whether one, two, or three points of bifurcation were measured. The branching pattern of the R vein varies tremendously among moths (Figs. 2 and 3), often but not always as a result of different numbers of total branches of this vein. Four measurements were taken for Sesiidae: *R* atrophies (seems to disappear because the vein walls disintegrated during the pupal stage) in sesiids at the most proximal points where R typically bifurcates, with the exceptions of two genera examined here, Bembecia Hübner and Melittia Hübner. In these genera, the cross-sectional diameter of R increases consistently until it first bifurcates, such that there is no optimal location for measuring this diameter. This same set of four measurements was also taken for Ctenuchina + Euchromiina because the first point of bifurcation in *R* was the only one that could be consistently measured from the photographs provided.

Hemolymph is known to leak out of the veins into the living tissue of the surrounding wing lamina [67], and the veins taper along the proximo-distal axis of the wing. To minimize the impact of vein tapering and leakage on the measurements, veins were measured as close as possible to the points where they bifurcate, as [68] have done. Here, all branches corresponding to the radius and radial sector (Rs) veins in Wootton's terminology [69] are treated as branches of R because all originate from the main R vein. A schematic of how we made our diameter and angle measurements is presented in Fig. 10.

All available wing slides from the Mississippi Entomological Museum and the Finnish Museum of Natural History were examined. The Lepidoptera slide drawers at the Smithsonian Institution that have been assigned United States National Museum numbers, which have the capacity to hold 155,548 slides, were searched for wing slides. All slides made with non-type material—i.e., slides made from specimens that have not been designated as name-bearers for their species-from the USA were taken on loan. This sampling method is unbiased in that all available material was examined, but the resulting dataset is taxonomically biased toward the taxa for which wing slides are typically made. Wing slides of zygaenoid moths housed in the California Department of Food and Agriculture were examined. Photographs of three wing slides of Agathiphaga vitiensis Dumbleton, taken for a recent study [26], were included. These photographs are of sufficient pixel density to measure the widths of veins and



were included because of the phylogenetic position of this taxon. Photographs of wing slides of Ctenuchina + Euchromiina were included because this taxon is not otherwise represented in the dataset. All of these wing slides were created for previous taxonomic and morphological studies [26, 70–82].

When these wing slides had been examined, the superfamilies Hepialoidea, Papilionoidea, and Bombycoidea were noticeably underrepresented in the dataset, with zero, one, and three wing slides, respectively. New wing slides for various species of Hepialidae were made from specimens at the EMEC (Essig Museum Entomological Collection, University of California, Berkeley) and the CASC (California Academy of Sciences). New wing slides for the hepialid species Gazoryctra mcglashani (Edwards) were made from two specimens provided by Laurence L. Crabtree, one of the two lepidopterists who recently documented re-encountering this species [83]. New wing slides for Papilionoidea and Bombycoidea were made from museum specimens from the EMEC and CASC and with specimens from the rainforest exhibit at the CAS that were collected after they perished. New wing slides for the atlas moth were made with specimens collected by Joshua Cluck at the Butterfly Habitat in Vallejo, California, after they perished.

These same measurements were also made from the wings of caddisflies (order Trichoptera), the sister group to Lepidoptera. The species examined were assigned to the subordinal clades identified by [84]. Rhyacophilidae, a family of controversial affinities, is the only representative in the dataset of the group traditionally referred to as Spicipalpia and now known as Integripalpia sensu [84]. In order to avoid confusion in the wake of recent changes to caddisfly taxonomy, this family is not discussed here as a representative of any higher clade. Only two measurements were made from each wing because the first point at which *R* bifurcates typically occurs very close to the base of the wing, where the edges of the veins are difficult to discern. Spread caddisflies from the EMEC and CASC were photographed for measurement at the EMEC.

Some caddisfly species, particularly belonging to the brevitentorian superfamily Leptoceroidea [84], have forewings with such a dense covering of hair that the veins can only be seen if the wings are removed and the hairs are brushed off—the same process used to create wing slides for Lepidoptera. One forewing was removed from three specimens each of *Phylloicus aeneus* (Hagen) and *Heteroplectron californicum* McLachlan (Calamoceratidae), and *Namamyia plutonis* Banks (Odontoceridae), at the EMEC. The wings broke apart when immersed in 70% ethanol, making it very difficult to remove the hairs before the wings became fragmented. Therefore, measurements of wing vein diameter could only be taken from two of the nine wings prepared.

We measured the diameters of vein cavities with photomicrographs that have a density of 146-4,660 pixels per millimeter depending on the size of the wing (mean value: 2,022 pixels/mm, 4,088,484 pixels/mm²). These photomicrographs were taken at Stanford University with a Leica M165 C microscope and a Leica DFC450 camera, at the Finnish Museum of Natural History (Luomus) with a Leica DM1000 LED and Leica S9D camera, and at the EMEC with a Leica S9i microscope and built-in camera. The slides were backlit with a stage light. With this optical setup, the boundary between the vein cavity and the vein wall is readily discernible. The widths were measured using the Pen Tool in Affinity Designer version 1.8.3, with the default Round Cap changed to the Butt Cap. The dimensions of each line were extracted from the Transform Panel in Affinity Designer and pasted into a Microsoft Excel spreadsheet.

Systematics

The representation of moth superfamilies among the specimens examined here roughly parallels the described species-level diversity of those superfamilies (Fig. 11). Gelechioidea, the most speciose superfamily of microlepidoptera [85], is noticeably overrepresented in the dataset. This superfamily is noted for its high proportion of undescribed species and may contain more true species-level diversity than any other superfamily in the order [86].



For the expository figure showing sampling effort, for the figures plotting various metrics against wing length, and for the linear regressions comparing these metrics to wing length, the paraphyletic grade Monotrysia, which contains the most early diverging superfamilies of extant Lepidoptera, was treated as a single superfamily. Although hepialid anatomy is exoporian rather than monotrysian [27], Hepialidae were included in the Monotrysia because they diverged within this grade, i.e., they diverged before the Ditrysia [85]. Monotrysia were lumped together because many of the monotrysian superfamilies contain a single family [85]. Because the superfamily Gracillarioidea is represented by fewer than 10 specimens in the dataset, and because this superfamily is suspected to be paraphyletic with respect to Yponomeutoidea [87], Gracillarioidea, and Yponomeutoidea were combined into a "Yponomeutoidea s.l." taxon. Because no explicitly phylogenetic analyses were conducted for this study, and because all analyses of individual superfamilies were conducted separately, these decisions to lump superfamilies did not impact any of the results.

The subtribes Ctenuchina and Euchromiina belong to the superfamily Noctuoidea [88]. However, because Ctenuchina and Euchromiina are unique among Noctuoidea in that they often have clear wings, and because they are sister taxa that form a monophyletic clade [89], we treat them as a separate group here. "Noctuoidea" is used here to describe all noctuoid moths except for Ctenuchina + Euchromiina.

Figures and analyses

For the main scatterplot illustrating the values of k found in this study, each point represents the mean value for 20 bifurcations (before the data were binned into groups of 20 bifurcations, they were arranged by the variable of interest, either d_0 or angle). This decision was made because plots of the raw data (i.e., values of d_0 , bifurcation angle, and k for individual bifurcations) are difficult to comprehend—not only because of the size of the dataset, but because the wide range of values of k precludes differentiation between k = 2, k = 3, etc. (Additional file 2: Figure S3). The decision to plot mean values for groups of 20 bifurcations vastly reduced the range of the data, permitting inferences at a much finer scale. Of the 338 bifurcations in which one child vein is wider than the parent vein and one is narrower than the parent vein, three have values of k that are so negative that they cannot be calculated with the software used (R version 4.0.2), and thus were removed from the dataset.

Four datasets are presented in the main scatterplot, with different kinds of processing. The first dataset is unprocessed, with nothing truncated or removed. In the second dataset, asymmetrical bifurcations (in which one child branch is wider than the parent and the other is narrower) were removed. In the third dataset, asymmetrical bifurcations were removed and values of k were truncated at -14 and 20. In the fourth dataset, asymmetrical bifurcations were removed, bifurcations in which both child branches are wider than the parent were removed, and k was truncated at 20.

Analyses were performed on the raw data rather than the truncated or binned data. To evaluate the d_0 threshold at which conformity to Murray's Law changes, change point analysis was performed with the mcp package version 0.3.0 [90] in R version 4.0.2 [91] with the default settings (Gaussian family, 9000 iterations from 3 chains). A generalized linear model for the effects of vein diameter and microscope magnification was performed on the data collected at the Finnish Museum of Natural History (Luomus), with magnification coded as a quantitative variable represented by the number of pixels corresponding to one millimeter.

Supplementary Information

The online version contains supplementary material available at https://doi.org/10.1186/s12915-021-01130-0.

Additional file 1: Supplement 1. Supplementary methods, results, and discussion of the potential impact of image quality. **Figure S1.** Values of *k* for bifurcations measured at the Finnish Museum of Natural History (Luomus), categorized by the magnification of the microscope when the wing slides were photographed. **Figure S2.** Measurement error, in

microns, of the diameters of 100 wing veins that were measured twice at with the same microscope at the same magnification.

Additional file 2: Figure S3. The raw relationship among d₀ and k.

Additional file 3: Supplement 2. Supplementary discussion of phylogenetic context.

Additional file 4: Figure S4. The raw relationship among bifurcation angle and *k*.

Additional file 5: Supplement 3. Supplementary discussion of generalization of Murray's Law for insect wings under flow transport, structural function, and total vein network mass constraints.

Acknowledgements

Various individuals assisted with the procurement of specimens for the creation of wing slides: Peter T. Oboyski, Roberta Brett, and Casey Hubble at the Essig Museum of Entomology, University of California Berkeley; Christopher Grinter, David Bettman, Tim Wong, and Freeland Dunker at the California Academy of Sciences; Joshua Cluck of the Butterfly Habitat in Vallejo, California; and Lawrence Crabtree. Becky Simmons of the University of North Dakota provided photographs of wing slides of Ctenuchina and Euchromiina. Many individuals provided assistance locating wing slides at various institutions, as follows. Mississippi Entomological Museum; M. Alma Solis, Conrad C. Labandeira, Floyd Shockley, Paul Z. Goldstein, Mark Metz, S. Augusta Maccracken. California Department of Food and Agriculture: Marc E. Epstein. Finnish Museum of Natural History, Luomus: Lauri Kaila, Pasi Sihvonen, Maria Heikkilä. Andrés Baresch and two anonymous reviewers provided valuable discussion of the ideas presented here.

Authors' contributions

SRS, CKB, and DL conceptualized the study. SRS, JLP, and DL designed the analyses. SRS prepared all specimens, collected all of the data, wrote the code, and drafted the initial version of the manuscript. DL drafted the "Generalization of Murray's Law" section of the supplement. SRS, JLP, CKB, and DL and edited the manuscript. The authors read and approved the final manuscript.

Funding

S.R.S. received funding from the Coleman F. Fung Interdisciplinary Graduate Fellowship, Vice Provost of Graduate Education, Stanford University, from the Harriet Benson Fellowship Award, Department of Geological Sciences, Stanford University, and from the McGee/Levorsen Research Grant, Department of Geological Sciences, Stanford University. None of these funders had any role in the design of the study, in the collection, analysis, and interpretation of data, or in writing the manuscript.

Availability of data and materials

All data and code are available at https://purl.stanford.edu/wq177gw5815.

Declarations

Ethics approval and consent to participate Not applicable.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

Author details

¹ Department of Geological Sciences, Stanford University, Stanford, USA. ² Department of Mechanical Engineering, Stanford University, Stanford, USA. ³ Faculty of Science and Engineering, University of Groningen, Groningen, The Netherlands.

Received: 28 December 2020 Accepted: 20 August 2021 Published online: 15 September 2021

References

 Le Guyader H. Geoffroy Saint-Hilaire: A Visionary Naturalist. Chicago: University of Chicago Press; 2004, p. 302.

- 2. Grimaldi D, Engel M. Evolution of the Insects. New York: Cambridge University Press; 2005, p. 772.
- 3. Dudley R. The Biomechanics of Insect Flight. Princeton N.J.: Princeton University Press; 2000, p. 496.
- Pass G. Beyond aerodynamics: The critical roles of the circulatory and tracheal systems in maintaining insect wing functionality. Arthropod Struct Dev. 2018;47(4):391–407.
- West G, Brown J. The origin of allometric scaling laws in biology from genomes to ecosystems: Towards a quantitative unifying theory of biological structure and organization. J Exp Biol. 2005;208(Pt 9):1575–92.
- 6. LaBarbera M. Principles of design of fluid transport systems in zoology. Science. 1990;249(4972):992–1000.
- LaBarbera M. The Design of Fluid Transport Systems: A Comparative Perspective. In: Bevan J, Kaley G, Rubanyi G, editors. Flow-Dependent Regulation of Vascular Function. New York, NY: Springer; 1995. p. 3–27.
- da Vinci L. The Notebooks of Leonardo Da Vinci. Oxford: Oxford University Press; 1980.
- Murray C. The physiological principle of minimum work. I, The vascular system and the cost of blood volume. Proc Natl Acad Sci. 1926;12:207–14.
- Marcinek D, LaBarbera M. Quantitative branching geometry of the vascular system of the Blue Crab, *Callinectes sapidus* (Arthropoda, Crustacea): A test of Murray's Law in an open circulatory system. Biol Bull. 1994;186(1):124–33.
- Sherman T. On connecting large vessels to small. The meaning of Murray's law. J Gen Physiol. 1981;78(4):431–53.
- 12. Cassot F, Lauwers F, Lorthois S, Puwanarajah P, Duvernoy H. Scaling laws for branching vessels of human cerebral cortex. Microcirculation. 2009;16(4):331–44.
- Cassot F, Lauwers F, Lorthois S, Puwanarajah P, Cances-Lauwers V, Duvernoy H. Branching patterns for arterioles and venules of the human cerebral cortex. Brain Res. 2010;1313:62–78.
- McCulloh K, Sperry J, Adler F. Murray's law and the hydraulic vs mechanical functioning of wood. Funct Ecol. 2004;18(6):931–8.
- Price C, Knox S-J, Brodribb T. The influence of branch order on optimal leaf vein geometries: Murray's Law and area preserving branching. PLOS ONE. 2013;8(12):85420.
- Wasserthal L. Antagonism between hemolymph transport and tracheal ventilation in an insect wing (*Attacus altas* L.): A disproof of the generalized model of insect wing circulation. J Comp Physiol B. 1981;147:27–40.
- Tsai C-C, Childers R, Nan Shi N, Ren C, Pelaez J, Bernard G, Pierce N, Yu N. Physical and behavioral adaptations to prevent overheating of the living wings of butterflies. Nat Commun. 2020;11(1):551.
- Wasserthal L. 7. Respiratory system. In: Kristensen N, editor. Lepidoptera: Moths and Butterflies 2 Handbuch Der Zoologie/Handbook of Zoology. Berlin & New York: Walter de Gruyter; 2003. p. 189–204.
- Appel E, Heepe L, Lin C-P, Gorb S. Ultrastructure of dragonfly wing veins: Composite structure of fibrous material supplemented by resilin. J Anat. 2015;227(4):561–82.
- Thomas M, Walsh K, Wolf M, McPheron B, Marden J. Molecular phylogenetic analysis of evolutionary trends in stonefly wing structure and locomotor behavior. Proc Natl Acad Sci. 2000;97(24):13178–83.
- Bergmann P, Richter S, Glöckner N, Betz O. Morphology of hindwing veins in the shield bug *Graphosoma italicum* (Heteroptera: Pentatomidae). Arthropod Struct Dev. 2018;47(4):375–90.
- Schieber G, Born L, Bergmann P, Körner A, Mader A, Saffarian S, Betz O, Milwich M, Gresser G, Knippers J. Hindwings of insects as concept generator for hingeless foldable shading systems. Bioinspiration Biomimetics. 2017;13(1):016012.
- 23. Zeller H. Blut und fettkörper im flügel der mehlmotte *Ephestia kühniella* Zeller. Z Morphol Okol Tiere. 1938;34(5):663–738.
- 24. Wootton R. Functional morphology of insect wings. Ann Rev Entomol. 1992;37:113–40.
- Stephenson D, Patronis A, Holland D, Lockerby D. Generalizing Murray's law: An optimization principle for fluidic networks of arbitrary shape and scale. J Appl Phys. 2015;118(17):174302.
- Schachat S, Gibbs G. Variable wing venation in *Agathiphaga* (Lepidoptera: Agathiphagidae) is key to understanding the evolution of basal moths. R Soc Open Sci. 2016;3(10):160453.
- Kristensen N. 5. The Homoneurous Glossata. In: Kristensen N, editor. Lepidoptera: Moths and Butterflies 1 Handbuch Der Zoologie/Handbook of Zoology. Berlin & New York: Walter de Gruyter; 1998. p. 51–63.

- Davis D. 6. The Monotrysian Heteroneura. In: Kristensen N, editor. Lepidoptera: Moths and Butterflies 1 Handbuch Der Zoologie/Handbook of Zoology. Berlin & New York: Walter de Gruyter; 1998. p. 65–90.
- Davis D, Robinson G. 7. The Tineoidea and Gracillarioidea. In: Kristensen N, editor. Lepidoptera: Moths and Butterflies 1 Handbuch Der Zoologie/Handbook of Zoology. Berlin & New York: Walter de Gruyter; 1998. p. 91–117.
- Dugdale J, Kristensen N, Robinson G, Scoble M. 8. The Yponomeutoidea. In: Kristensen N, editor. Lepidoptera: Moths and Butterflies 1 Handbuch Der Zoologie/Handbook of Zoology. Berlin & New York: Walter de Gruyter; 1998. p. 119–30.
- Nielsen E, Common I. Lepidoptera (moths and butterflies). In: Naumann I, editor. The Insects of Australia: A Textbook for Students and Research Workers. Carlton: Melbourne University Press; 1991. p. 817–915.
- 32. Baixeras J. An overview of genus-level taxonomic problems surrounding *Argyroploce* Hübner (Lepidoptera: Tortricidae), with description of a new species. Ann Entomol Soc Am. 2002;95(4):422–31.
- Eichlin T, Duckworth W, Vol. 5. Sesioidea: Sesiidae. Washington DC: Wedge Entomological Research; 1988, p. 176.
- Epstein M, Geertsema H, Naumann C, Tarmann G. 10. The Zygaenoidea. In: Kristensen N, editor. Lepidoptera: Moths and Butterflies 1 Handbuch Der Zoologie/Handbook of Zoology. Berlin & New York: Walter de Gruyter; 1998. p. 159–80.
- Ackery P, De Jong R, Vane-Wright R. 16. The butterflies: Hedyloidea, Hesperioidea and Papilionoidea. In: Kristensen N, editor. Lepidoptera: Moths and Butterflies 1 Handbuch Der Zoologie/Handbook of Zoology. Berlin & New York: Walter de Gruyter; 1999. p. 263–300.
- Hodges R. 9. The Gelechioidea. In: Kristensen N, editor. Lepidoptera: Moths and Butterflies 1 Handbuch Der Zoologie/Handbook of Zoology. Berlin & New York: Walter de Gruyter; 1998. p. 131–58.
- Munroe E, Solis M. 14. The Pyraloidea. In: Kristensen N, editor. Lepidoptera: Moths and Butterflies 1 Handbuch Der Zoologie/Handbook of Zoology. Berlin & New York: Walter de Gruyter; 1998. p. 233–56.
- Schachat S, Goldstein P. Acronictinae (Lepidoptera: Macroheterocera: Noctuidae) demonstrate the variable role of wing venation in the evolution of the nymphalid groundplan. Insect Syst Divers. 2018;2(2):1–15.
- Schachat S. The wing pattern of *Hydriomena* Hübner, [1825] (Lepidoptera: Geometridae: Larentiinae) lacks a predictable relationship with venation. J Morphol. 2019;280(11):1651–67.
- Lemaire C, Minet J. 18. The Bombycoidea and their relatives. In: Kristensen N, editor. Lepidoptera: Moths and Butterflies 1 Handbuch Der Zoologie/Handbook of Zoology. Berlin & New York: Walter de Gruyter; 1998. p. 322–53.
- Revellin R, Rousset F, Baud D, Bonjour J. Extension of Murray's law using a non-Newtonian model of blood flow. Theor Biol Med Model. 2009;6(1):7.
- Boppana S, Hillyer J. Hemolymph circulation in insect sensory appendages: Functional mechanics of antennal accessory pulsatile organs (auxiliary hearts) in the mosquito *Anopheles gambiae*. J Exp Biol. 2014;217(17):3006–14.
- 43. Chintapalli R, Hillyer J. Hemolymph circulation in insect flight appendages: Physiology of the wing heart and circulatory flow in the wings of the mosquito, *Anopheles gambiae*. J Exp Biol. 2016;219(24): 3945–51.
- Nielsen E, Kristensen N. The Australian moth family Lophocoronidae and the basal phylogeny of the Lepidoptera – Glossata. Invertebr Syst. 1996;10(6):1199.
- 45. Wasserthal L. Heartbeat reversal and its coordination with accessory pulsatile organs and abdominal movements in lepidoptera. Experientia. 1976;32(5):577–9.
- Fristrom D, Gotwals P, Eaton S, Kornberg T, Sturtevant M, Fristrom J. Bistered: A gene required for vein/intervein formation in wings of Drosophila. Development. 1994;120:2661–71.
- 47. Coates M, Clack J. Polydactyly in the earliest known tetrapod limbs. Nature. 1990;347(6288):66–9.
- Alberch P, Gale E. Size dependence during the development of the amphibian foot. Colchicine-induced digital loss and reduction. Development. 1983;76(1):177–97.
- Alberch P, Gale E. A developmental analysis of an evolutionary trend: Digital reduction in amphibians. Evolution. 1985;39(1):8–23.
- 50. Hanken J. A Morphological novelty in the limb skeleton accompanies miniaturization in salamanders. Science. 1985;229(4716):871–4.

- de Sá R, Tonini J, van Huss H, Long A, Cuddy T, Forlani M, Peloso P, Zaher H, Haddad C. Multiple connections between Amazonia and Atlantic Forest shaped the phylogenetic and morphological diversity of *Chiasmocleis* Mehely, 1904 (Anura: Microhylidae: Gastrophryninae). Mol Phylogenet Evol. 2019;130:198–210.
- Scherz M, Hutter C, Rakotoarison A, Riemann J, Rödel MO, Ndriantsoa S, Glos J, Roberts S, Crottini A, Vences M, Glaw F. Morphological and ecological convergence at the lower size limit for vertebrates highlighted by five new miniaturised microhylid frog species from three different Madagascan genera. PLOS ONE. 2019;14(3):0213314.
- Wirkner C, Tögel M, Pass G. The Arthropod Circulatory System. In: Minelli A, Boxshall G, Fusco G, editors. Arthropod Biology and Evolution: Molecules, Development, Morphology. Berlin, Heidelberg: Springer; 2013. p. 343–391.
- 54. Braun A. Pupal tracheation and imaginal venation in Microlepidoptera. Trans Am Entomol Soc. 1933;59(3):229–68.
- 55. Braun A. Observations on the pupal wings of *Nepticula*, with comparative notes on other genera. Ann Entomol Soc Am. 1917;10(3):233–9.
- Kristensen N. Respiratory system of the primitive moth *Micropterix* calthella (Linnaeus) (Lepidoptera : Micropterigidae). Int J Insect Morphol Embryol. 1984;13(2):137–56.
- 57. Williams H, Trask R, Weaver P, Bond I. Minimum mass vascular networks in multifunctional materials. J R Soc Interface. 2008;5(18):55–65.
- Hoskins P. The Arterial System II, Forces, Adaptability and Mechanotransduction. In: Hoskins P, Lawford P, Doyle B, editors. Cardiovascular Biomechanics. Cham: Springer International Publishing; 2017. p. 83–106.
- Huo Y, Kassab G. Intraspecific scaling laws of vascular trees. J R Soc Interface. 2012;9(66):190–200.
- 60. West G, Brown J, Enquist B. A general model for the origin of allometric scaling laws in biology. Science. 2013;276(5309):122–6.
- Zamir M, Sinclair P, Wonnacott T. Relation between diameter and flow in major branches of the arch of the aorta. J Biomech. 1992;25(11):1303–10.
- Kristensen N. 4. The Non-Glossatan Moths. In: Kristensen N, editor. Lepidoptera: Moths and Butterflies 1 Handbuch Der Zoologie/Handbook of Zoology. Berlin & New York: Walter de Gruyter; 1998. p. 41–9.
- 63. Davis D. A monograph of the family Arrhenophanidae (Lepidoptera: Tineoidea). Smithson Contrib Zool. 2003;620:1–80.
- Kitching I, Rawlins J. 19. The Noctuoidea. In: Kristensen N, editor. Lepidoptera: Moths and Butterflies 1 Handbuch Der Zoologie/Handbook of Zoology. Berlin & New York: Walter de Gruyter; 1998. p. 355–401.
- Minet J, Scoble M. 17. The Drepanoid / Geometroid Assemblage. In: Kristensen N, editor. Lepidoptera: Moths and Butterflies 1 Handbuch Der Zoologie/Handbook of Zoology. Berlin & New York: Walter de Gruyter; 1998. p. 301–20.
- 66. Arnold J. Blood circulation in insect wings. Mem Entomol Soc Can. 1964;96(S38):5–60.
- Wasserthal L. Haemolymph flows in the wings of Pierid butterflies visualized by vital staining (insecta, lepidoptera). Zoomorphology. 1983;103:177–92.
- Carvalho M, Turgeon R, Owens T, Niklas K. The hydraulic architecture of Ginkgo leaves. Am J Bot. 2017;104(9):1285–98.
- 69. Wootton R. Function, homology and terminology in insect wings. Syst Entomol. 1979;4:81–93.
- Albrecht A, Kaila L. Variation of wing venation in Elachistidae (Lepidoptera: Gelechioidea): Methodology and implications to systematics. Syst Entomol. 1997;22(3):185–98.
- DaCosta M, Weller S. Phylogeny and classification of Callimorphini (Lepidoptera: Arctiidae: Arctiinae). Zootaxa. 2005;1025(1):1–94.
- DaCosta M, Larson P, Donahue J, Weller S. Phylogeny of milkweed tussocks (Arctiidae: Arctiinae: Phaegopterini) and its implications for evolution of ultrasound communication. Ann Entomol Soc Am. 2006;99(5):723–42.
- Heikkilä M, Mutanen M, Kekkonen M, Kaila L. Morphology reinforces proposed molecular phylogenetic affinities: A revised classification for Gelechioidea (Lepidoptera). Cladistics. 2013;30:563–89.
- Jaeger C. Phylogeny of Tortricidae (Lepidoptera): A morphological approach with enhanced whole mount staining techniques. M.S. Thesis. Mississippi State University. 2017.
- 75. Kaila L. Phylogeny and classification of the Elachistidae s.s, (Lepidoptera: Gelechioidea). Syst Entomol. 1999;24(2):139–69.

- Kaila L. Phylogeny of the superfamily Gelechioidea (Lepidoptera: Ditrysia): An exemplar approach. Cladistics. 2004;20(4):303–40.
- Kaila L, Vol. 11. Elachistine Moths of Australia (Lepidoptera: Gelechioidea: Elachistidae) Monographs on Australian Lepidoptera. Melbourne: CSIRO Publishing; 2011.
- Lee S, Brown R. Phylogenetic relationships of Holarctic Teleiodini (Lepidoptera: Gelechiidae) based on analysis of morphological and molecular data. Syst Entomol. 2008;33(4):595–612.
- Lee S, Brown R. Review of Symmocinae (Lepidoptera: Autostichidae) in North America with the description of a new genus and species. J Lepidopterists' Soc. 2010;64(4):177–87.
- Mabey J. Phylogeny of the Tiger Moth genus *Phoenicoprocta* Druce (Noctuidae: Arctiinae: Euchromiina) for use in examining the evolution of courtship systems. M.S. Thesis. University of North Dakota. 2010.
- Sihvonen P, Kaila L. Phylogeny and tribal classification of Sterrhinae with emphasis on delimiting Scopulini (Lepidoptera: Geometridae). Syst Entomol. 2004;29(3):324–58.
- Sihvonen P. Phylogeny and classification of the Scopulini moths (Lepidoptera: Geometridae, Sterrhinae). Zool J Linnean Soc. 2005;143(4): 473–530.
- Wagner D, Crabtree L. Rediscovery of *Gazoryctra mcglashani* in montane meadows of Middle Sierra Nevada Mountains (Lepidoptera: Hepialidae). J Lepidopterists' Soc. 2018;72(3):241–3.
- Thomas J, Frandsen P, Prendini E, Zhou X, Holzenthal R. A multigene phylogeny and timeline for Trichoptera (Insecta). Syst Entomol. 2020;45(3):670–86.
- 85. Nieukerken E, Kaila L, Kitching I, Kristensen N, Lees D, Minet J, Mitter C, Mutanen M, Regier J, Simonsen T, Wahlberg N, Yen S-H, Zahiri R, Adamski D, Baixeras J, Bartsch D, Bengtsson BA, Brown J, Bucheli S, Davis D, de Prins J, De Prins W, Epstein M, Gentili-Poole P, Gielis C, Hättenschwiler P, Hausmann A, Holloway J, Kallies A, Karsholt O, Kawahara A, Koster S, Kozlov M, Lafontaine J, Lamas G, Landry J-F, Lee S, Nuss M, Park K-T, Penz C, Rota J, Schintlmeister A, Schmidt B, Sohn J-C, Solis M, Tarmann G, Warren A, Weller S, Yakovlev R, Zolotuhin V, Zwick A. Order Lepidoptera Linnaeus, 1758. In: Zhang Z-Q, editor. Animal Biodiversity: An Outline of Higher-Level Classification and Survey of Taxonomic Richness. Auckland: Zootaxa; 2011. p. 212–21.
- Mitter C, Davis D, Cummings M. Phylogeny and evolution of Lepidoptera. Ann Rev Entomol. 2017;62(1):265–83.
- Sohn J-C, Regier J, Mitter C, Davis D, Landry J-F, Zwick A, Cummings M. A molecular phylogeny for Yponomeutoidea (Insecta, Lepidoptera, Ditrysia) and its implications for classification, biogeography and the evolution of host plant use. PLoS One. 2013;8(1):55066.
- Zahiri R, Holloway J, Kitching I, Lafontaine J, Mutanen M, Wahlberg N. Molecular phylogenetics of Erebidae (Lepidoptera, Noctuoidea). Syst Entomol. 2012;37(1):102–24.
- Dowdy N, Keating S, Lemmon A, Lemmon E, Conner W, Chialvo C, Weller S, Simmons R, Sisson M, Zaspel J. A deeper meaning for shallow-level phylogenomic studies: Nested anchored hybrid enrichment offers great promise for resolving the tiger moth tree of life (Lepidoptera: Erebidae: Arctiinae). Syst Entomol. 2020;45(4):874–93.
- Lindeløv J. Mcp: An R Package for Regression With Multiple Change Points. 2020.
- R Development Core Team. R: A Language and Environment for Statistical Computing. 2020.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

